# The Effect of Creatine on Immortalized Schwann Cell Proliferation

Peyton Kimmel, Caitlyn Henry, and Angela L. Asirvatham Department of Biology, Misericordia University, Dallas PA



### Introduction

Schwann cells support neurons in the peripheral nervous system. They function to myelinate the axons of neurons, which aids in the neuron's speed of conductivity. Schwann cells are also known to aid in the repair of neurons, when myelin is damaged<sup>1</sup>. Previous studies in Schwann cell cultures have shown that addition of heregulin, a neuronal growth factor, and forskolin, a pharmacological activator of cAMP, stimulates a synergistic growth response<sup>2</sup>. Although these growth factors and signaling molecules have been studied in Schwann cell growth, not much is known about creatine, an important component of the phosphocreatine energy buffer system that is crucial for providing ATP during neuronal repair<sup>3</sup>. Based on the significance of creatine in reducing neuronal losses<sup>4</sup>, we hypothesized that addition of creatine, with growth factors to Schwann cell cultures will stimulate proliferation. Therefore, the primary objective of this study is to determine the optimal dose and time point, at which creatine stimulates Schwann cell growth in cultures incubated with heregulin and forskolin.

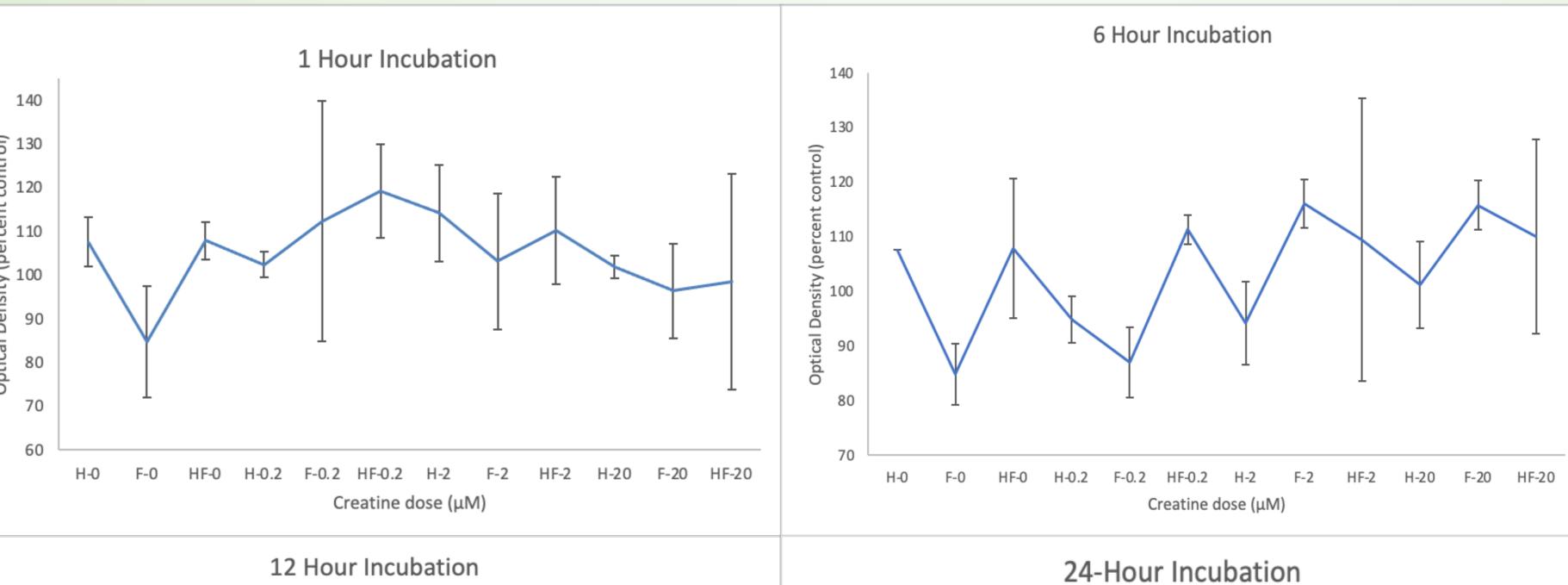
#### References

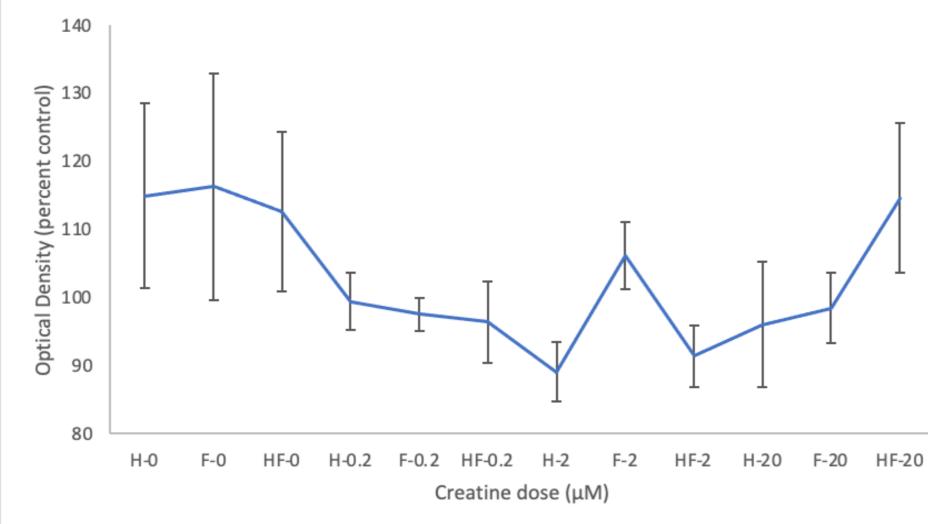
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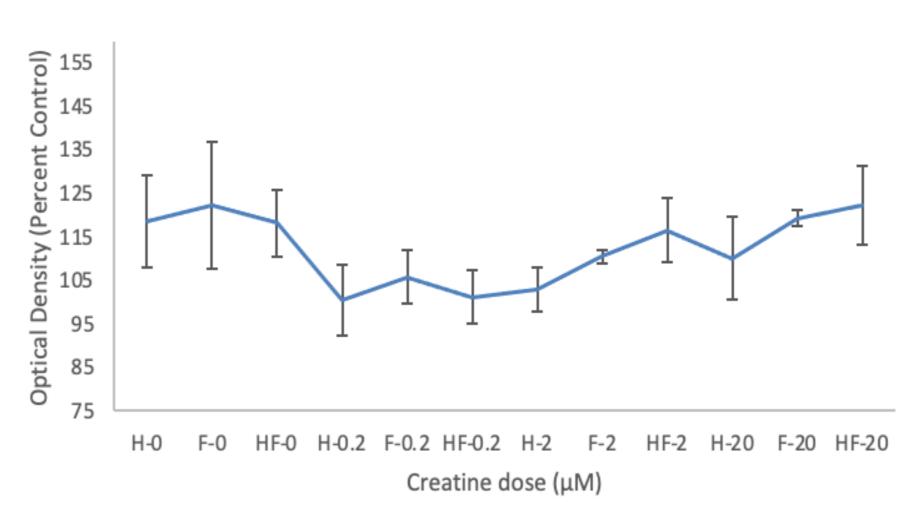
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## Creatine Dose Response







**Fig.1** – Effect of creatine on Schwann cell growth: Schwann cells from S16 cell line (SC-2941, ATCC, Manassas, VA) were incubated with N2 (control medium) (0), heregulin (H), forskolin (F), or heregulin + Forskolin (HF) for 1, 6, 12, or 24 hours along with 0,  $0.2\mu m$ ,  $2\mu M$  or  $20\mu M$  creatine. To determine cell growth, a colorimetric proliferation assay using MTT (3-[4,5-dimethylthiazol-2-yl]-2,5,-diphenyltetrazolium bromide) (Thermofisher Scientific, Waltham, MA) was performed. MTT was added to the cells for four hours followed by incubation with SDS. Optical density was measured using the Spectramax plate reader (Molecular Devices, San Jose, CA) and analyzed with SoftmaxPro software. The experiment was replicated three times. Using the SPSS software package, data was analyzed by ANOVA. Post hoc tests for comparison between means were further analyzed using Least Significant Difference. A p value < 0.05 was considered statistically significant (Table 1)

## Conclusions

#### 1-Hour

• Addition of  $0.2\mu M$  and  $2\mu M$  creatine increased proliferation for all treatments in comparison to control.

#### 6-Hour

• Addition of  $0.2\mu M$  creatine stimulated cell growth in control and heregulin+forskolin treated cultures. Addition of  $2\mu M$  CR increased growth in cells treated with heregulin and forskolin in comparison to control

#### 12-Hour

• A dose of  $0.2\mu M$  creatine increased proliferation in control cultures. Forskolinstimulated cells elicited the highest proliferation at a dose of  $0\mu m$  of creatine in comparison to control

#### 24-Hour

• Cells treated with both heregulin and forskolin displayed highest proliferation at a dose of  $2\mu M$  creatine. Cells incubated with  $0.2\mu M$  and creatine showed a reduction in proliferation for all treatments in comparison to control.

In summary, forskolin increased proliferation at all time points implying that creatine stimulates cAMP-mediated pathways at the  $2\mu M$  concentration. Addition of both heregulin and forskolin for 24 hours with  $2\mu M$  creatine resulted in a substantial increase in proliferation. These results suggest that creatine-induced proliferation of Schwann cells appear to be influenced by dose and incubation period.

## P-values for different dose and time of Creatine Treatment

Table 1

#### Dose and Growth Factor p-value 0μM CR F, 2μM CR F 0.029 0.03 0μM CR F, 20μM CR F 0.043 0.2μM CR F, 20μM CR F 0.027 12 hour 0μM CR H, 2μM CR H 0μM CR H, 2μM CR HF 0.042 0.02 0μM CR F, 2μM CR H 0μM CR F, 2μM CR HF 0.032 0μM CR HF, 2μM CR H 0.042 2μM CR HF, 20μM CR HF 0.045 24 hour OμM CR N2, OμM CR F 0.03 0μM CR N2, 20μM CR HF 0.031 0μM CR F, 0.2μM CR N2 0.03 0μM CR F, 0.2μM CR H 0.033 0μM CR F, 0.2μM CR HF 0.04 0μM CR F, 2μM CR N2 0.03 0μM CR F, 20μM CR N2 0.03 0.2μM CR N2, 20μM CR HF 0.031 0.2μM CR H, 20μM CR HF 0.034 0.2μM CR HF, 20μM CR HF 0.04 0.031 2μM CR N2, 20μM CR HF 20μM CR N2, 20μM CR HF 0.031